

Plastic Responses of Sweet Potato (*Ipomeabatatas L.*) Root Anatomy in Compacted Soil Conditions

Mamuda Umaru Ali

*Department of Crop Science, Universiti Putra Malaysia, Selangor, Malaysia;
Department of Agriculture Hassan Usman Katsina Polytechnic, Katsina, Nigeria*

SakiminSitiZaharah

- 1. Department of Crop Science, Faculty of Agriculture, Universiti Putra Malaysia, 43400 UPM Serdang Selangor*
- 2. Institute of Tropical Agriculture & Food Security (ITAFoS), Universiti Putra Malaysia, 43400 UPM Serdang, Selangor, Malaysia, Seri Kembangan, Selangor,*

Adam Puteh

Department of Crop Science, Faculty of Agriculture, Universiti Putra Malaysia, 43400 UPM Serdang Selangor

Daljit Singh Karam Singh

Department of Land Management, Faculty of Agriculture, Universiti Putra Malaysia, 43400 UPM Serdang Selangor

Muhammad HazimNazli

Department of Crop Science, Faculty of Agriculture, Universiti Putra Malaysia, 43400 UPM Serdang Selangor

Abstract-The response of crops to abiotic stress largely depends on its root system architecture since the root is the first to sense abiotic stress prevailing in the soil media. This study was conducted to investigate the effect of soil compaction on the anatomy of sweet potato roots. A field experiment was conducted in March 2017. The treatments used consisted of three compaction levels (no-till, tilled once and tilled twice) and three varieties (Gendut, Kedudut, and Vitato). The treatments were in factorial combination; arranged in a split-plot design and replicated four times. Sampled roots were viewed under a binocular microscope (Olympus BCH) for anatomical studies. The collected data were subjected to analysis of variance (ANOVA) to determine the effect of the treatments and their interactions; using the general linear model; SAS software version 9.4. Differences among treatment means were compared using the least significant difference (LSD) at $p \leq 0.05$. Results obtained, indicated cortex and root xylem diameters, root proliferation was decreased with increased compaction but root lignification was increased. Root structure was also influenced by varietal variation. However, the effect was not constant with varieties. In all the observed characters Kedudut and Gendut had the least and the highest mean values respectively. Gendut variety showed the highest ability to adapt to compacted soil. The decrease in anatomical features could lead to a decrease in yield as a result of possible restrictions in the uptake of soil water and nutrients.

Keywords - Root anatomy, Root development, Soil compaction, Sweet potato.

I. INTRODUCTION

The root is a plant organ that differs in structure and functions; and is found in close contact with growth media [1]. Root ensures plant productivity through the uptake of soil resources. Growing roots are capable of sensing and responding to the particular stimulus which is in most cases induced by physical, chemical, and biological factors in the soil media [2]. The resultant response varies with time, environment, and genotype. The resource use efficiency of most crops including sweet potato could be improved through the understanding of root development[3]. Root varies greatly

between species and between individuals of any particular genotypes [2] [4] and has high plasticity response to environmental conditions such as soil compaction, nutrient, water, and oxygen availability [5] [6] [7] [8] [9]. Sweet potato root consists of structural features. These features show great modifications when encountered with environmental stress. This will lead to reduced water and nutrient uptake [10]. As the result, the plant nutrient utilization efficiency is reduced and consequent to this; is the lowering of the plant productivity.

Soil compaction is a structural degradation of the soils where the sizes and shapes of the soil pores are altered thereby leading to a decrease in soil macropores and an increase in both soil bulk density and soil strength [11] [3]. Root growth, development, and extension are inhibited in compacted soils [12]. Thereby resulting in poor root penetration and the emergence of a shallow root system. Plants when confronted with hard surfaces; also have to expend more energy for root growth [13], thereby; the available energy needed for other growth and developmental processes becomes altered. Consequent to these effects, moisture, and nutrients might be in short supply and thus the plants become stressed and stunted.

Compacted soils with high strength may limit root growth through a large mechanical resistance to root growth and development. The roots in turn respond to mechanical impedance by slowing down the process of root extension per given time and increasing the diameter rates at a location behind the root tip [14]. In potato, the roots were found restricted at a shallower depth and were unable to penetrate deeply with an increased degree of soil compaction [15]. The production of sweet potato (*Ipomoea batatas*) covers all major regions of the world (tropics and subtropics). Sweet potato is important in the human diet in terms of its contribution to body nutritional requirements. It also contributes to industrial sustainability and in livestock production. It ranks 5th as the most important food crop after wheat, rice maize, and cassava [16] and 2nd after cassava among root and tuber crops [17]. Malaysia is ranked 56th among sweet potato producers worldwide this is because the crop is considered as a minor in the country, however, it is one of the popular cash crops grown by the small-scale peasant farmers for the fresh market [18]. There exist over 20 different varieties of sweet potato in Malaysia [19] both local and improved.

Root growth and performance have been extensively studied in: maize [20] [11] [21], sugarcane [21] barley [24] and carrot [25] [26]. However, studies related to this issue are scarce in sweet potato [27]. [28] and [29] provided statistical evidence showing none existence of literature relating to sweet potato root anatomy. The few studies so far conducted on sweet potato roots were mostly done in pots. For a more realistic approach, there is a need to carry out such experiments on the field; to depict the real situation prevailing within the soil environment. The usage of heavy agricultural implements and chemical fertilizers in farm operations coupled with heavy rainfall and the practice of continuous cropping makes the soil in peninsular Malaysia prone to degradation and increased compaction and hence the need to study this environmental menace. It is therefore the objective of this study was to investigate the anatomical responses of sweet potato roots, under compacted soil conditions.

II. MATERIALS AND METHODS

A field experiment was conducted at Ladang 15; Teaching and Research Farm of University Putra Malaysia (UPM). The site is approximately located at **2°99'13.16"N** and **101°73'34.55"E** and 32m above sea level. The climate of the area is equatorial, being hot and humid throughout the year. The average annual temperature, relative humidity and rainfall are: 27°C, 90%, and 250cm respectively. The prevailing wind patterns are southeast monsoon and the northeast monsoon that blow from May-September and November-March and these influence light and heavy rainfall in the area respectively [30].

Before the setup of the experiment, soil samples were taken randomly from the experimental plots, with a tubular auger at the depth of 0-30cm. The collected samples were; bulked, air-dried, sieved (using 2 mm mesh), and analyzed for composite (type) and physicochemical properties using the standard procedure as described by [31]. Penetration resistance and soil moisture content at 10 cm depth were measured using a chart recording Eijkelkamp Stiboka penetrometer with a 1-cm² cone ('Cone 1' 60° top angle, and 8-mm-diameter shaft) used by the manufacturer's (Eijkelkamp Agriscerch Equipment, 6987 EM Giesbee, Netherland) recommendations. The penetrometer was inserted to 30 cm.

Three sweet potato varieties (Gendut, Vitato, and Kedudut) that varied genetically and are popularly produced amongst the sweet potato farmers in the study area were used. The treatments tested were in factorial combination of three levels of soil compaction (untilled = NT, tilled once = T1 and tilled twice = T2) and three varieties (Gendut, Vitato, and Kedudut). The treatments were arranged in a split-plot design with tillage as the main plot and variety in subplot and

replicated four times. In each of the experimental plots (15m²), three plants were randomly tagged and picked to record observations. Root samples were obtained destructively by the excavation of soil cores using Shovelomics technic as described by [32] and [33] using hand trowel and spades. After thorough washing of the sampled roots using a bucket, sieve mesh, and hosepipe; each plant sample was dissected into roots, stems, and leaves. The washed roots were then stored in a refrigerator at 3°C to reduce root respiration before the commencement of anatomical and morphological analysis.

Sampled roots for anatomical studies were placed in distilled water and sectioned. Transverse sections of about 15µm were done with a razor blade. The sectioned samples were stained with toluidine blue solution for about a minute as described by [34]. To reduce the excesses of the colorant, the samples were rinsed in water. The stained samples were then put on glass slides and viewed under a binocular microscope (Olympus BCH). Data on the following variables were observed and recorded: protoxylem element number, the diameter of the xylem vessels, the thickness of the cortex, and the presence of lignin.

The collected data were subjected to analysis of variance (ANOVA) to determine the effect of the treatments and their interactions; using the general linear model SAS software version 9.4. Differences among treatment means were compared using the least significant difference (LSD) at $p \leq 0.05$.

III. RESULTS AND DISCUSSION

3.1 Effect of tillage

Results on Table 1 showed that soil compaction had a significant effect on root xylem diameter, the thickness of the root cortex, and the number of lignified roots. However, it had no significant effect on the number of protoxylem elements having five or more xylem poles. The smallest xylem (4.74%) and root cortex diameter (0.68%) values were obtained from NT treatment. But, the highest number of lignified roots (42.93%) was obtained from NT treated plots. The largest xylem (39.42%) and root cortex diameter (27.82%) values and the least number of lignified roots (17.44) were, however, obtained with T1 treatment. NT treatment did not differ significantly with T2 treatments in all the observed parameters (Table 1). Correlation studies showed that; an increase in soil compaction index (bulk density) resulted in a significant increase (0.94) in the number of lignified roots. However, soil compaction had a negative but weak correlation with root xylem diameter and thickness of the root cortex (-0.15 and -0.18 respectively) (Table 2). The decrease in the sizes of these anatomical features may probably lead to a corresponding decrease in hydraulic conductivity of the xylem vessels and also, reduced carbohydrate storage capacity of the cortex. This may probably lead to; reduced plant growth as the result of hindrances in hydraulic conductivity and possibly, poor yield; owing to the low storage capacity of the cortex. In agreement with this finding, [35] reported a decrease in the cortex and root xylem diameter in more compacted soils when working with *Fraxinus Angustifolia* (Vahl) seedlings. An increase in soil compaction led to a significant increase in the number of lignified roots. The lignification process in root renders the root unable to develop into a storage root [36] and therefore, this is an indicator that the more the roots become lignified, the more the yield potential is reduced. The influence of soil compaction on the protoxylem element number was however not significant (Table 1). This trait could probably remain unaffected by an external stimulus; or possibly, the degree of compaction within the soil is quite insufficient to introduce changes, in growth and developmental pattern of protoxylem element number.

3.2 Effect of variety

Variety had a significant effect on root xylem diameter, the thickness of the root cortex, protoxylem element with five or more xylem pole numbers, and the number of lignified roots (Table 1). Gendut had the highest mean xylem diameter (31.33%), the thickness of the cortex (49.22%), and protoxylem element number (39%). However, it had the least number of lignified roots (29.17%). In contrast, Kedudut had the least means for xylem diameter (16.69%), the thickness of the root cortex (27.62%), and protoxylem element number (8.12) having five or more poles but it had the highest number (50.93%) of lignified roots. Vitato on the other hand differed significantly with both Gendut and Kedudut in all characters except the number of lignified roots: where it was at par with Kedudut. The largest mean diameters of root cortex and root xylem observed in the Gendut variety signified that Gendut may probably have better hydraulic conductivity and larger storage capacity and hence, may have greater yield potentials than Kedudut which had the least mean thickness of both cortex and xylem. Roots having five or more protoxylem element numbers are often associated with the possibilities of developing into storage roots [23] [36] and hence a variety (such as Gendut) having a higher number of roots with five or more protoxylem element is expected to give a higher yield than Kedudut. The lignification process in roots, tends to prevent the roots from developing into storage roots [36] and thus a variety

such as Kedudut that had a higher number of lignified roots may tend to promote lower yield production. This confirmed earlier findings [23] [36] that reported the significant influence of variety on protoxylem element number and presence and distribution of lignified roots in different varieties of the sweet potato crop.

Table 1. Effect of soil compaction and variety on: Root xylem diameter, Protoxylem element number, Thickness of the cortex, and Number of lignified roots on sweet potato at UPM

Treatment	Xylem diameter	Protoxylem element number	Thickness of the cortex	Number of lignified roots
Tillage (T)				
No-till (NT)	119.76b	2.58	152.34b	1.98a
Tilled once (T1)	196.35a	2.55	210.54a	1.13b
Tilled twice (T2)	18.95b	2.59	154.11b	1.81a
LSD	9.32	NS	1.78	0.44
P-value	≤0.0001	0.0082	≤0.0001	0.0305
Variety (V)				
Gendut	162.02a	3.00a	191.65a	1.58b
Kedudut	124.09c	1.83c	140.17c	3.24a
Vitato	148.92b	2.75b	185.17b	3.09a
LSD	5.54	0.21	1.75	0.54
P-value	≤0.0001	≤0.0001	≤0.0001	0.0424
T*V	1042.35	0.03	361.45	0.51
LSD	NS	NS	1.76	NS
P-value	≤0.0001	0.8551	≤0.0001	0.6463
CV (%)	4.45	16.59	1.18	11.23

Table 2. The correlation coefficient between soil compaction and anatomical characteristics of sweet potato root

	<i>SBD</i>	<i>RXD</i>	<i>TRCOT</i>	<i>LIG. ROOT</i>
SBD	1			
RXD	-0.11ns	1		
TRCOT	-0.15ns	0.99**	1	
LIG. ROOT	0.95**	0.20ns	0.15ns	1

Note: SBD = Soil bulk density, RXD = Root xylem diameter, TRCOT = Thickness of root cortex, LIG.ROOT = Lignified roots

3.3 Interaction effect between tillage and variety

Interaction between soil compaction and variety on the root cortex was also significant ($p=0.05$) (Table 1). All varieties showed thicker and thinner root cortexes with T1 and NT treatments respectively. The thickest root cortex was obtained from the combination of Gendut and T1 treatments. In contrast, Kedudut when combined with NT treatment produced the thinnest root cortexes. It is interesting to note that both Vitato and Kedudut at T1 treatment had larger root cortexes than Gendut under T2 treatment; even though, the variety Gendut had the thickest root cortex as shown on (Figure 1).

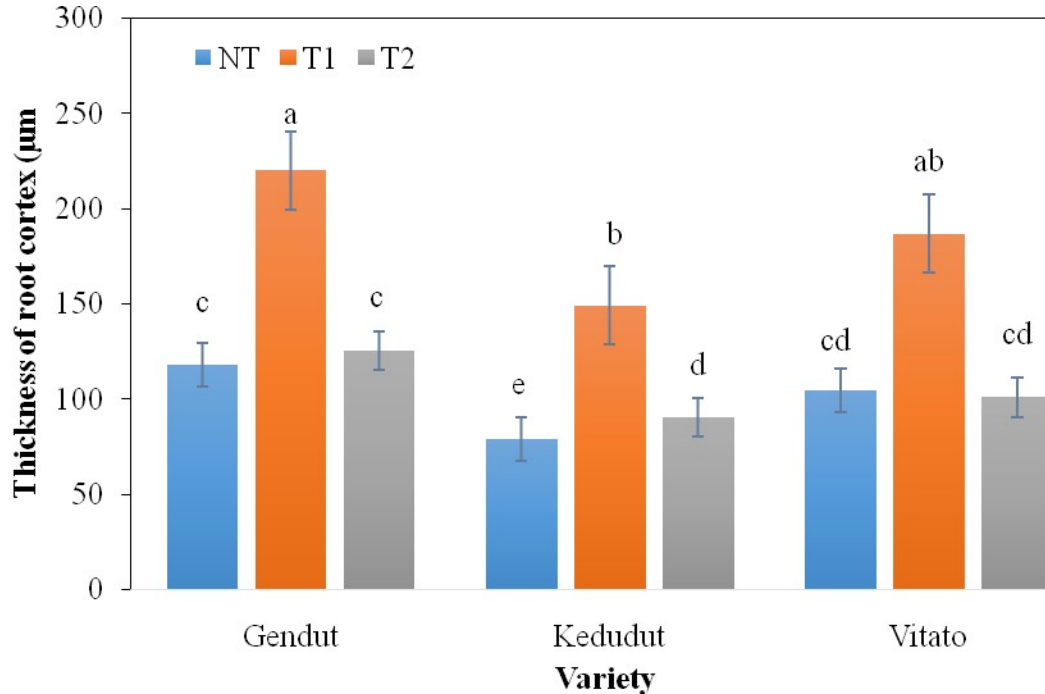


Figure 1. Interaction between soil compaction and variety on the thickness of sweet potato root cortex. Note: NT= No Tilled, T1 = Tilled Once and T2 = Tilled Twice.

IV. CONCLUSION / RECOMMENDATION

Environmental stress resulting from soil compaction adversely influenced the structural performance of sweet potato roots; by decreasing resource capture potentiality of the crop. However, the intensity of the effect varied with the type of variety and the degree of soil compaction. The more the soil becomes impended, the more the root traits underwent diminishing changes in growth patterns and development. In this study, however, moderately compacted soil (T1) tends to promote better root development than the high and less compacted treatments. Gendut variety had the highest potentials to adapt to compacted soils; having the highest mean xylem diameter, the thickness of the cortex, protoxylem element number, and the least number of lignified roots. A single tillage operation is adequate; to overcome soil compaction menace and Gendut exhibited promising traits for better performance in the study area. However, there is a need for further research to vindicate this assertion.

REFERENCES

- [1] Gregory, P.J. (2006). Roots, rhizosphere and soil: the route to a better understanding of soil science? *European Journal of Soil Science*, 57(1), 2-12.
- [2] Ristova, D., and Busch, W. (2014). Natural variation of root traits: from development to nutrient uptake. *Plant Physiology*, 166(2), 518-527.
- [3] Tracy, S.R., Black, C.R., Roberts, J. A., and Mooney, S.J. (2013). Exploring the interacting effect of soil texture and bulk density on root system development in tomato (*Solanum lycopersicum* L.). *Environmental and Experimental Botany*, 91, 38-47.
- [4] Gifford, M.L., Banta, J.A., Katari, M.S., Hulsmans, J., Chen, L., Ristova, D. and Birnbaum, K.D. (2013). Plasticity regulators modulate specific root traits in discrete nitrogen environments. *PLoS Genetics*, 9(9), e1003760.
- [5] Malamy, J.E. (2005). Intrinsic and environmental response pathways that regulate root system architecture. *Plant, Cell & Environment*, 28(1), 67-77.
- [6] Nibau, C., Gibbs, D.J., and Coates, J.C. (2008). Branching out in new directions: the control of root architecture by lateral root formation. *New Phytologist*, 179(3), 595-614.
- [7] Whitmore, A.P., and Whalley, W.R. (2009). Physical effects of soil drying on roots and crop growth. *Journal of Experimental Botany*, 60(10), 2845-2857.
- [8] Grubber, B. D., Giehl, R. F., Friedel, S., and von Wirén, N. (2013). Plasticity of the Arabidopsis root system under nutrient deficiencies. *Plant Physiology*, 163(1), 161-179.
- [9] Li, C., Yan, K., Tang, L., Jia, Z., & Li, Y. (2014). Change in deep soil microbial communities due to long-term fertilization. *Soil Biology and Biochemistry*, 75, 264-272.

- [10] Lipiec, J., Medvedev, V.V., Birkas, M., Dumitru, E., Lyndina, T.E., Rouseva, S., and Fulajtar, E. (2003). Effect of soil compaction on root growth and crop yield in Central and Eastern Europe. *International Agrophysics*, 17(2), 61-70.
- [11] Ramazan, M., Khan, G. D., Hanif, M., and Ali, S. (2012). Impact of soil compaction on root length and yield of corn (*Zea mays*) under irrigated condition. *Middle-East Journal of Scientific Research*, 11(3), 382-385.
- [12] Agbede, T.M., and Adekiya, A.O. (2011). Evaluation of Sweet Potato (*Ipomoea Batatas* L.) Performance and Soil properties under Tillage Methods and Poultry Manure Levels. *Emirates Journal of Food and Agriculture*, 164-177.
- [13] Daniel, M.S. (2010). Effect of plastic mulches and row cover on yield and quality of sweet potato. A thesis submitted to graduate faculty of Auburn University in: Agbede, O. A and Adekiya, G. O (2011). Evaluation of Sweet potato Performance and Soil Properties under Tillage methods and Poultry manure. *Environmental Journal of Food and Agriculture* 2011:23(2):164-177.
- [14] Malamy, J. E. (2005). Intrinsic and environmental response pathways that regulate root system architecture. *Plant, Cell & Environment*, 28(1), 67-77.
- [15] Ariel, F., Diet, A., Verdenaud, M., Gruber, V., Frugier, F., Chan, R., and Crespi, M. (2010). Environmental regulation of lateral root emergence in *Medicago truncatula* requires the HD-Zip 1 transcription factor HB1. *The Plant Cell*, 22(7), 2171-2183.
- [16] Mukhopadhyay, S.K., Chattopadhyay, A., Chakraborty, I., and Bhattacharya, I. (2011). Crops that feed the world 5. Sweetpotato. Sweetpotatoes for income and food security. *Food Security*, 3(3), 283.
- [17] FAO (2014) FAO statistical databases FAOSTAT. <http://faostat3.fao.org/> Accessed 8 November 2014
- [18] Tan, S.L., Aziz, A.A., and Zaharah, A. (2007). Selection of sweet potato clones for flour production. *Journal of Tropical Agriculture and Food Science*, 35(2), 205-212.
- [19] Zaharah, A., and Tan, S.L. (2006). Performance of selected sweet potato varieties under different growing seasons on bris sandy soil in Malaysia. In *14th Triennial Symposium of International Society of Tropical Root Crops* (pp. 20-26).
- [20] Imhoff, S., Kay, B.D., Da Silva, A.P., and Hajabbasi, M.A. (2010). Evaluating responses of maize (*Zea mays* L.) to soil physical conditions using a boundary line approach. *Soil and Tillage Research*, 106(2), 303-310.
- [21] Otto, R., Silva, A.D., Franco, H.C.J., Oliveira, E.D., and Trivelin, P.C.O. (2011). High soil penetration resistance reduces sugarcane root system development. *Soil and Tillage Research*, 117, 201-210.
- [22] Huang, G.B., Qiang, C.H.A.I., Feng, F.X., and Yu, A.Z. (2012). Effects of different tillage systems on soil properties, root growth, grain yield, and water use efficiency of winter wheat (*Triticum aestivum* L.) in arid Northwest China. *Journal of Integrative Agriculture*, 11(8), 1286-1296.
- [23] Beleh, T., Hammes, P.S., and Robbertse, P. (2004). The origin and structure of adventitious roots in sweet potato (*Ipomoea batatas*). *Australian Journal of Botany*, 52(4), 551-558.
- [24] Bingham, I.J., and Bengough, A.G. (2003). Morphological plasticity of wheat and barley roots in response to spatial variation in soil strength. *Plant and Soil*, 250(2), 273-282.
- [25] Rosenfeld, A.B. (1997). Effects of nitrogen and soil conditions on growth, development and yield in potatoes. PhD thesis, University of Cambridge.
- [26] Kristensen, H.L., and Thorup-Kristensen, K. (2007). Effects of vertical distribution of soil inorganic nitrogen on root growth and subsequent nitrogen uptake by field vegetable crops. *Soil Use and Management*, 23(4), 338-347.
- [27] Villordon, A., LaBonte, D., Solis, J., and Firon, N. (2012). Characterization of lateral root development at the onset of storage root initiation in 'Beauregard' sweetpotato adventitious roots. *HortScience*, 47(7), 961-968.
- [28] Villordon, A.Q., Ginzberg, I., and Firon, N. (2014). Root architecture and root and tuber crop productivity. *Trends in Plant Science*, 19(7), 419-425.
- [29] Khan, M.A., Gemenet, D.C., and Villordon, A. (2016). Root system architecture and abiotic stress tolerance: current knowledge in root and tuber crops. *Frontiers in plant science*, 7, 1584.
- [30] MMD (2017). Malaysian Meteorology Department. Ministry of Science, Technology and Innovation. Kuala Lumpur, Malaysia. www.met.gov.my weather record for Pusat Pertanian Serdang (2013-2019).
- [31] Gregorich, E.G., and Carter, M.R. (2007). *Soil sampling and methods of analysis*. CRC press.
- [32] Trachsel, S., Kaeppler, S.M., Brown, K.M., and Lynch, J.P. (2011). Shovelomics: high throughput phenotyping of maize (*Zea mays* L.) root architecture in the field. *Plant and Soil*, 341(1-2), 75-87.
- [33] Smith, S., and De Smet, I. (2012). Root system architecture: insights from Arabidopsis and cereal crops.
- [34] Yeung, E.C. (1998). A beginner's guide to the study of plant structure. *Tested studies for laboratory teaching*, 19, 125-141.
- [35] Alameda, D., and Villar, R. (2012). Linking root traits to plant physiology and growth in *Fraxinus angustifolia* Vahl. Seedlings under soil compaction conditions. *Environmental and Experimental Botany*, 79, 49-57.
- [36] Villordon, A.Q., La Bonte, D.R., Firon, N., Kfir, Y., Pressman, E., and Schwartz, A. (2009). Characterization of adventitious root development in sweetpotato. *HortScience*, 44(3), 651-655.